

Research Article

Phytochemical Profiling and Anticancer Effect of *Fagonia arabica* on MCF-7 Breast Cancer Cells

Ambreen Sher¹, Sobia Tabassum^{1*}, Fikri Nashrullah², Aysha Sarfraz Rizvi³, Farhana Amin⁴, Sania Begum⁵, Farkhanda Jabeen⁵,

¹Department of Biological Sciences, Faculty of Sciences, International Islamic University, Islamabad, Pakistan.

²Master of Fisheries Science Department, Faculty of Agriculture, Gadjah Mada University, Yogyakarta, Indonesia

³School of Science, Minzu University of China, Beijing China.

⁴National Institute for Plant Genomics and Biotechnology (NIGAB), National Agriculture Research Council, Islamabad, Park Road, Islamabad 45500, Pakistan.

⁵Department of Zoology, Wildlife and Fisheries, Arid Agriculture University, Rawalpindi,

*Corresponding author's email: sobia.tabasum@iiu.edu.pk

Abstract

This research analyzes the phytochemicals in *Fagonia arabica* leaves, revealing a strong presence of flavonoids, terpenoids, saponins, anthraquinones, cardiac glycosides, and tannins. The methanolic fraction, rich in gallic acid and rutin equivalencies, exhibited exceptional antioxidant activity, validated through 2,2-diphenyl-1-picrylhydrazyl (DPPH) and phosphomolybdate assays. Employing FTIR and GCMS spectra, we objectively assessed the diverse chemical constituents, emphasizing the credibility of this method in identifying mix-substance systems in traditional and herbal medicine. To further understand the therapeutic potential, MCF-7 cells were treated and cytotoxicity assays (MTT and acid phosphatase assay) were conducted to get IC₅₀ values that further confirmed the anticancer properties of *Fagonia arabica*. Real-time PCR analysis targeted Caspases and Wnt/ β -catenin genes, revealing statistically significant (***) results with upregulated caspases and downregulated Wnt signaling genes. The findings highlight *Fagonia arabica* extract as a highly effective therapeutic strategy against breast cancer cells, strategically modulating Wnt/ β -catenin genes without causing significant damage to normal cells. This study shows potential in identifying novel bioactive chemicals in these medicinal plants, which will help us better grasp their therapeutic potential.

Keywords: *Fagonia arabica*, Wnt signaling, Phytochemical, Breast cancer cell lines.

Article History: Received: 14 Apr 2025, Revised: 16 Apr 2025, Accepted: 24 Apr 2025, Published: 30 Apr 2025.

Creative Commons License: NUST Journal of Natural Sciences (NJNS) is licensed under Creative Commons Attribution 4.0 International License.



Introduction

Embryonic development relies on signaling pathways. Mutations in these pathways in adults can cause abnormal cell growth. The Wnt pathway, crucial for tissue development and cell regulation, involves Wnt ligands binding to Frizzled receptors, triggering biological responses [1, 2, 3].

Breast cancer stands as a significant cause of morbidity and mortality globally among women. Advanced methods are being developed to detect primary tumors, metastases, and recurrent disease for effective management. Research indicates that dysregulation of Wnt/ β -catenin signaling significantly fuels tumorigenesis, compromising patient survival and treatment efficacy [4]. While chemotherapy can impede tumor growth, prolonged use of single agents often yields poor results. Combination therapies are now standard, yet systemic drug delivery causes widespread damage to healthy cells, leading to adverse effects [5]. Recent studies uncover molecular pathways associated with breast cancer proliferation, metabolism, and resistance to targeted therapies [1]. Understanding these mechanisms is vital to combat innate and acquired drug resistance and enhance treatment outcomes.

Natural products and medicinal plants have long been integral to healthcare. About half of current medications are derived from natural sources, highlighting their importance. Understanding how these natural chemicals are absorbed and distributed in the body is critical [6]. Resveratrol, Curcumin, and Quercetin are a few plant-derived substances that may block the Wnt signaling pathway, which is essential for cancer development [7,8]. While salinomycin, albeit synthetic, targets cancer stem cells by blocking the Wnt pathway and shows promise for cancer treatment. Similarly, Curcumin, found in

turmeric, has demonstrated anticancer capabilities in preclinical trials [9, 10]. They are still being tested for safety and efficacy in humans, emphasizing the need for thorough evaluation in cancer treatment. Given the side effects and medicine resistance associated with chemotherapy, the high rate of breast cancer in Pakistan is very concerning. The current study compares natural and herbal treatments with conventional pharmaceuticals to see which is more effective against breast cancer cell lines. One such comparison examines the possible anticancer effects of *Fagonia arabica*. Widely found across the Indian subcontinent, *Fagonia* species are tropical herbs of the Zygophyllaceae family. A major component in Ayurvedic medicine, *Fagonia arabica* is mostly used for its antihypertensive [11], anti-inflammatory [12], analgesic, and thrombolytic properties [13]. Its medicinal use depends on identifying Dhamasa via morphological and microscopic analysis [14]. For its health-promoting qualities, *Fagonia arabica* is used as a component of herbal beverages and infusions. Despite not being a common food source, folk medicine occasionally uses it as a dietary supplement to promote general health [15]. Being valued for its therapeutic qualities, and some of its parts are eaten or used as herbal remedies for their health advantages. *Fagonia arabica*, which is rich in alkaloids and flavonoids [16], may help the body combat free radicals and reduce oxidative stress [17]. This research aims to assess the effectiveness of *Fagonia arabica* targeting Wnt/ β -catenin signaling genes in MCF-7 cells, in light of its potential anticancer properties. Despite the lack of clinically approved medications that specifically target Wnt/ β -catenin signaling in cancer, there is a lot of interest in this field because several approaches are being researched.

Materials and Methods

Sample preparation

The specimens of *Fagonia arabica* collected from various locations in Pakistan have the voucher number PMAS-346. Arid Agriculture University Rawalpindi plant taxonomist Prof. Dr. Rahmatullah Qureshi recognized the specimens. The plant material was cleaned, dried in the shade for three weeks, and then ground into a powder. To guarantee a uniform particle size, it was sieved through a 60-mesh topology Willy mill, and the powder was packed within an airtight container. The preparation of the *Fagonia arabica* leaf extract (FALE) involved the use of the maceration technique [18].

Phytochemical analysis

The phytochemical analysis of the *Fagonia arabica* leaf extract (FALE) was carried out using the methodology of [19], which may have medical applications.

Antioxidant assays

To evaluate the antioxidant capability against various kinds of free radicals, two antioxidant assays were conducted.

Measuring DPPH radical scavenging activity

The assay involved serial dilution of standards and plant extract, followed by incubation in the dark with a 0.004 % DPPH stock solution with 517 nm absorption measurement [20].

TAC assay

The assay involved preparing plant extract stock solutions in DMSO, creating diluted samples, and combining them with a reagent solution, determining the total antioxidant capacity using the standard curve [19].

TPC / TFC contents

The total phenolic content of FALE was determined using gallic acid equivalents, or milligrams of gallic acid per gram of dried extract. The total flavonoid contents (mg rutin/ g dry extract) were estimated using rutin equivalents. The blank was generated from methanol using the methods given by [18].

Fourier Transform Infrared Spectrophotometer (FTIR)

FTIR spectroscopic analysis of the FALE identified many functional molecules with distinct peak values. The material was crushed in a 1:10 ratio with KBr to prepare the samples, and a hydraulic press was used to shape the material into pellets. The FTIR Spectroscopy device, which has a 400–4000 cm^{-1} scanning array and a 4 cm^{-1} resolution, was then used to analyze these pellets [21].

Gas chromatography mass spectrometry analysis

The method [21] was used to analyze the volatile components of FALE. By comparing retention periods with spectrum data from the Wiley and NIST libraries, the validity of the chemicals found by GCMS analysis was verified [22].

Cell culturing

Using the human breast cancer cell line MCF-7 (from ECACC), the study was carried out with ethical approval number IIUI/ORIC/Bioethics/110-81. The cells were cultivated grown and supplemented, according to standard growth conditions [23].

Cytotoxicity assay

MCF-7 cells were fed with Curcumin (10 mM stock) as a reference drug to evaluate FALE effects. The range of FALE administration was 0–500 $\mu\text{g/ml}$. The

methodology described by [20] was followed for both the MTT and Acid Phosphatase assays.

Selection of doses

Plant cytotoxicity tests were used to choose the doses, and they were then modified to roughly match their IC₅₀ values. Before being processed for real-time PCR expression, MCF-7 cells received two doses of treatment.

RNA extraction and expression profiling of targeted

RNA was extracted from the treated MCF-7 cells following the method described by [24]. Subsequently, using a cDNA synthesis kit, the RNA was converted into cDNA. The expression of genes, including Caspase-1, -3, -7, -9, Wnt-3a, and β -catenin, was assessed using the cDNA as a template. The housekeeping gene GAPDH was used as a control. Expression profiling was conducted by qPCR in triplicates for all treated and untreated samples, employing the SYBR green method. The $2^{-\Delta\Delta CT}$ technique of [25], was used to assess relative expression. Lists of the primers and the ideal PCR conditions are given in supplementary Tables.

Statistical analysis

All analyses were done in triplicate using mean \pm SEM, and one one-way ANOVA was applied using Graph Pad Prism where required. Given the qualitative nature of analysis for FTIR and GCMS statistical analysis was not applicable.

Results

Percentage yield of extracts

The methanolic extract of *Fagonia arabica* yielded 18g per 100g dry powder.

Phytochemical analysis

The FALE exhibited the presence of alkaloids, phenols, flavonoids, terpenoids, coumarins, glycosides, quinones, and saponins, as shown in Table 1.

Table 1: Phytochemical analysis of selected plant extract.

Sr. no.	Phytochemical Test	FA
1	Alkaloids	+
2	Flavonoids	+
3	Phenols	+
4	Tannins	+
5	Terpenoids	+
6	Saponins	+
7	Carboxylic Acid	+
8	Cardiac Glycosides	+
9	Coumarins	+
10	Oxylates	+
11	Sterols	+

FALE= *Fagonia arabica* Leaves extract (+) = Presence of Phytochemical, (-) = Absence of phytochemical.

Antioxidant assay

The following antioxidant assays were performed to screen free radical scavengers.

DPPH Free radicals assay

The DPPH assay revealed the antioxidant potential of FALE, with results presented in Table 2. The EC₅₀ value for FALE was determined to be 0.51 ± 0.02 mg/ml, and dose-related activity was observed (Figure 1A). A significant correlation among all doses of the plant was detected through one-way ANOVA.

Phosphomolybdenum (Total Antioxidant capacity) assay

The phosphomolybdate assay was employed to evaluate antioxidant capacity of FALE which was measured in terms of

ascorbic acid equivalency (AEE), with results illustrated in Figure 1B and summarized in Table 3. A direct relationship was observed between the total antioxidant capacity and FALE concentration, with a decrease in AEE value with each serial dilution. Significant differences among different concentrations were determined via one-way ANOVA ($P < 0.0001$, $R^2 = 0.9368$).

TPC/ TFC

The total phenolic content of FALE was found to be 78.72 ± 0.5 mg gallic acid/ g of extract, while the total flavonoid content was measured at 50.02 ± 0.1 mg rutin/ g of extract, as illustrated in Figure 1C, 1D, and Table 4.

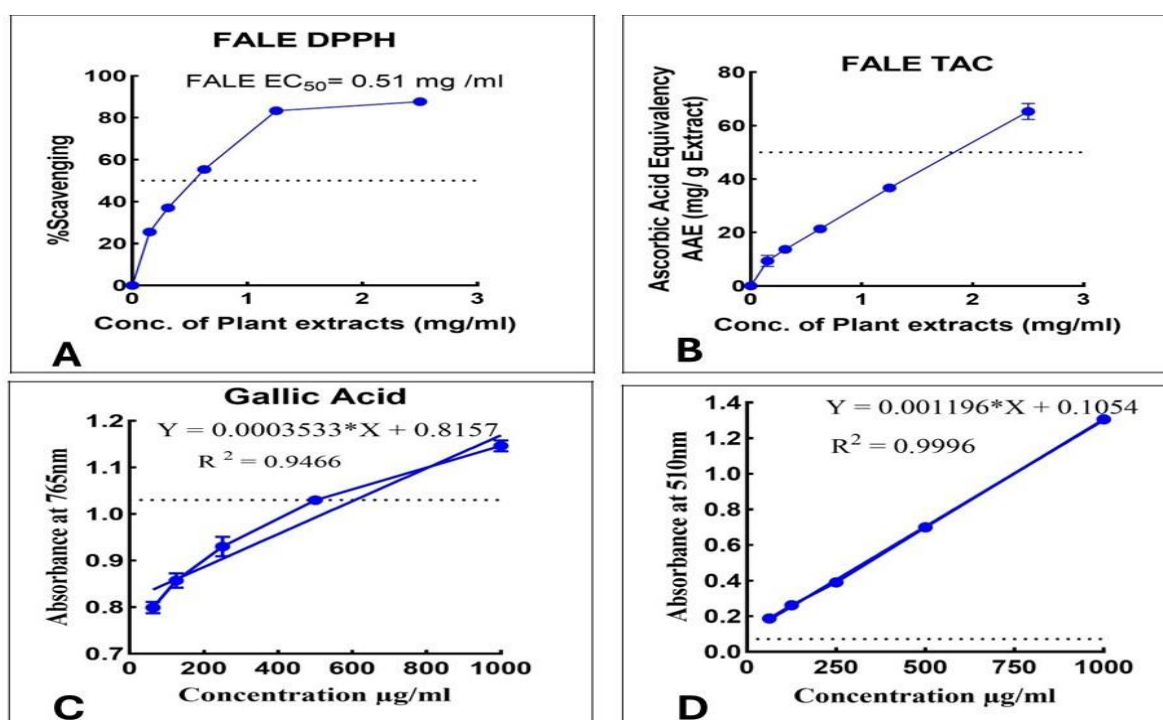


Figure 1: **A** Graphical representation of DPPH free radical scavenging activity of FALE at different concentrations. **B.** Graphical representation Total antioxidant Capacity assay for FALE at different concentrations. **C.** Graphical representations of Total Phenolic Contents of FALE using Gallic acid as a standard. **D** Total Flavonoid Contents of FALE using Rutin as a standard. Each value represents a mean \pm SE ($n = 3$).

Table 2: Percentage scavenging (mean \pm SEM) Values for *Fagonia arabica* at different concentrations.

Conc. of Samples mg/ mL	A. A	FALE
2.5	92 \pm 1	88 \pm 0
1.25	92 \pm 0	84 \pm 1
0.62	92 \pm 0	56 \pm 1
0.31	92 \pm 0	34 \pm 4
0.16	92 \pm 1	29 \pm 5

Values are presented as Mean \pm SEM ($n = 3$), the percentage of DPPH free radical scavenging by A.A (Ascorbic Acid) and the methanolic extracts of FALE (*Fagonia arabica* Leaves extract).

Table 3: Phosphomolybdate assay showing ascorbic acid equivalency (AEE) mg/ g of *Fagonia arabica* extract (mean \pm SEM).

Conc. of FALE (mg/ml)	Ascorbic Acid Equivalency (AEE) mg/g of <i>Fagonia arabica</i> Extract (mean \pm SEM)
2.5	65 \pm 2
1.25	36 \pm 2
0.62	21 \pm 1
0.31	13 \pm 1
0.15	9 \pm 1

Values are presented as Mean \pm SEM (n = 3). Results of Phosphomolybdenum assay by the Methanolic plant extracts of *Fagonia arabica*.

Table 4: Quantitative equivalencies of FALE contents with standards.

Sample	TPC (GAE mg/ g extract)	TFC (RE mg/ g extract)
FALE	78.72 \pm 0.5	50.02 \pm 0.1

GAE= Gallic acid equivalence, RE= Rutin equivalence.

FTIR analysis

Our samples confirmed the presence of plants secondary metabolites through FTIR analysis, revealing functional groups present in plant extracts. Notably, peaks at specific wave numbers indicated various functional groups. For instance, the peak at 3020 cm^{-1} indicated the stretching vibration of OH, while the signal at 1709 cm^{-1} suggested HC=O stretching vibration, indicating the presence of carbonyl compounds (aldehydes). Changes in the intensity ratio of bands at 1536 cm^{-1} for CH stretching and the presence of aliphatic compounds at 1430 cm^{-1} were observed.

Additionally, peaks at 1302 cm^{-1} (C=O) indicated the presence of ketones, and 1173 cm^{-1} (C-H) signified alkanes. Furthermore, the peak at 881 cm^{-1} represented the peak of primary alcohol glucose. These FTIR results outlined the spectrum of medicinally

essential plants that are beneficial for applications in the agricultural industry (Table 5, Figure 2).

Table 5: FTIR analysis showing wave numbers of functional groups identified in for FALE.

Plant extract	Peak Values cm^{-1}	Functional group	Reference
FALE	3020	H-C=O stretching	[48]
	1536	(C-H stretching) alkanes	[49]
	1302	Normal aliphatic esters	[50]
	1173	C=O stretching	[51]
	882	-OH Primary alcohol	[52]

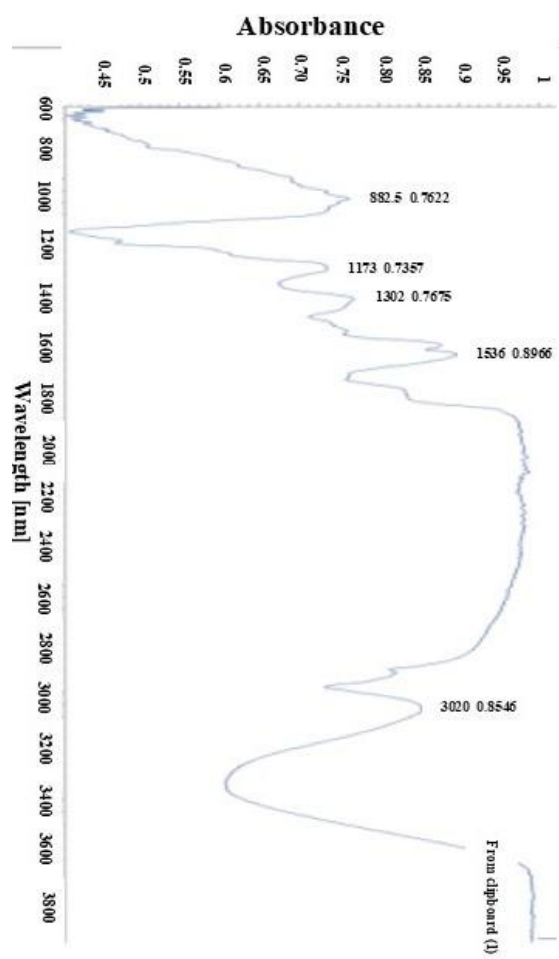


Figure 2: FTIR analysis showing functional groups of important phytochemicals present in FALE.

GC-MS analysis

GC-MS chromatograms of FALE illustrated various retention times, facilitating the identification of bioactive compounds present. By comparing mass spectra with the NIST library, 23 bioactive compounds were characterized. These compounds may contribute to the therapeutic benefits associated with the plant. Notably, 2,3-Diphenyl-5-methoxybenzoic-1,4 (58.78% abundance) and 3,7,11,15-Tetramethyl-2-hexadec (11.81% abundance) were identified as the most abundant polyphenols in FALE (Figure 3, Table 6).

Cell culturing

MCF-7 and MDA-MB-231 breast cancer cell lines were initially cultured for the study. However, due to mycobacterial contamination in MDA-MB-231, the study proceeded only with MCF-7 cell lines.

Cytotoxicity assays

To assess the cytotoxic effects of essential phytochemicals, present in FALE, MTT, and Acid Phosphatase (APT) assays were conducted. As per previous studies, a dose range of 0 to 500 $\mu\text{g/ml}$ was utilized for the cytotoxicity assays.

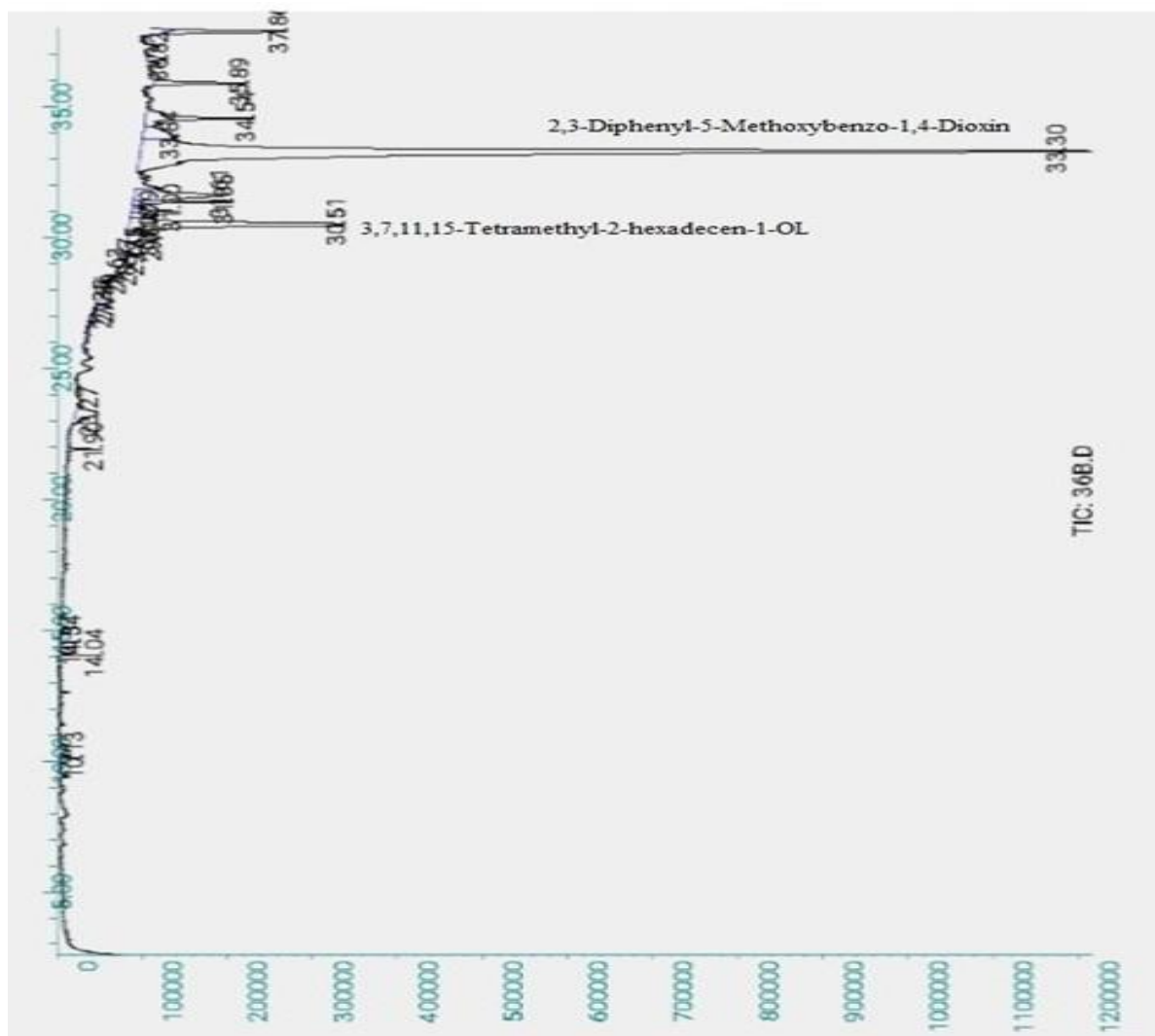
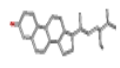
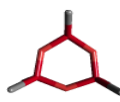
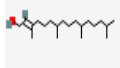
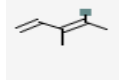
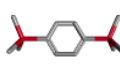
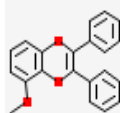
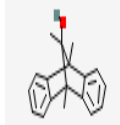
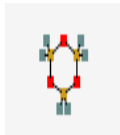
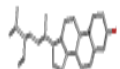
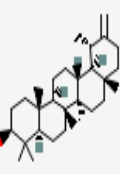


Figure 3: GCMS analysis Showing peaks for the presence of volatile components present in FALE.

Table 6: GCMS analysis of *F. arabica* showing bioactive compounds.

RT	Compound name	CAS No.	Abundance	Mol. formula	Mol. Weight	Compound Structure	Bioactivity	Ref
23.27	Stigmast-4-en-3-one	001058-61-383	2.04	C ₂₉ H ₄₈ O	412.7		Hypoglycemic	[52]
29.94	Hexamethyl Cyclo-trisiloxane	000541-05-943	1.36	C ₆ H ₁₈ O ₃ Si ₃	222.46		antioxidant and antidiabetic activities	[53]
30.52	3,7,11,15-Tetramethyl-2-hexadecen-1-OL	102608-53-7	11.81	C ₂₀ H ₄₀ O	296.5		Anti-cancerous, anti microbial, antidiuretic anti oxidant	[53]
31.35	Stigmastan-3,5-diene	1000214-16-4	2.17	C ₆ H ₁₀	82.14		Antimicrobial	[54]
31.61	Silane, 1,4-phenylenebis	013183-70-5	5.66	C ₁₂ H ₂₂ Si ₂	222.47		Antibacterial, anti larvacidal, antiviral, anti cancerous	[55]
33.30	2,3-Diphenyl-5-Methoxybenzo-1,4-Dioxin	091201-56-8	58.78	C ₂₁ H ₁₆ O ₃	316.3		Anticancerous,	[56]
33.84	9,10-Methanoanthracen-11-Ol	084783-02-8	2.69	C ₁₈ H ₁₈ O	250.3		Antitumor	[57]
34.54	Cyclotrisiloxane	000541-05-9	4.44	H ₆ O ₃ Si ₃	138.3		antimicrobial	[58]
35.89	β-Sitosterol	000083-46-5	2.10	C ₂₉ H ₅₀ O	414.7		Anti-inflammatory antipyretic	[59]
37.87	Taraxasterol	001059-14-9	2.99	C ₃₀ H ₅₀ O	426.7		Anti-inflammatory, chemo-preventive	[60]

MTT and APT assay

The impact of FALE on MCF-7 cells was evaluated using MTT and acid phosphatase assays. The dosage was determined based on the initial antioxidant analysis, ranging from 0 to 500 $\mu\text{g/ml}$. After incubating the cells with test samples for 48 hours, the percentage inhibition results were determined. Curcumin was used as a standard natural drug to compare with the

samples under study via MTT and APT assays. The IC₅₀ values for all samples are presented in Table 7 and Figure 4.

Table 7: Cytotoxicity analysis of FALE

Sample	MTT assay IC ₅₀ ($\mu\text{g/ml}$)	APP assay IC ₅₀ ($\mu\text{g/ml}$)
FALE	181 $\mu\text{g/ml}$	109 $\mu\text{g/ml}$
Curcumin	13.7 $\mu\text{g/ml}$	11.5 $\mu\text{g/ml}$

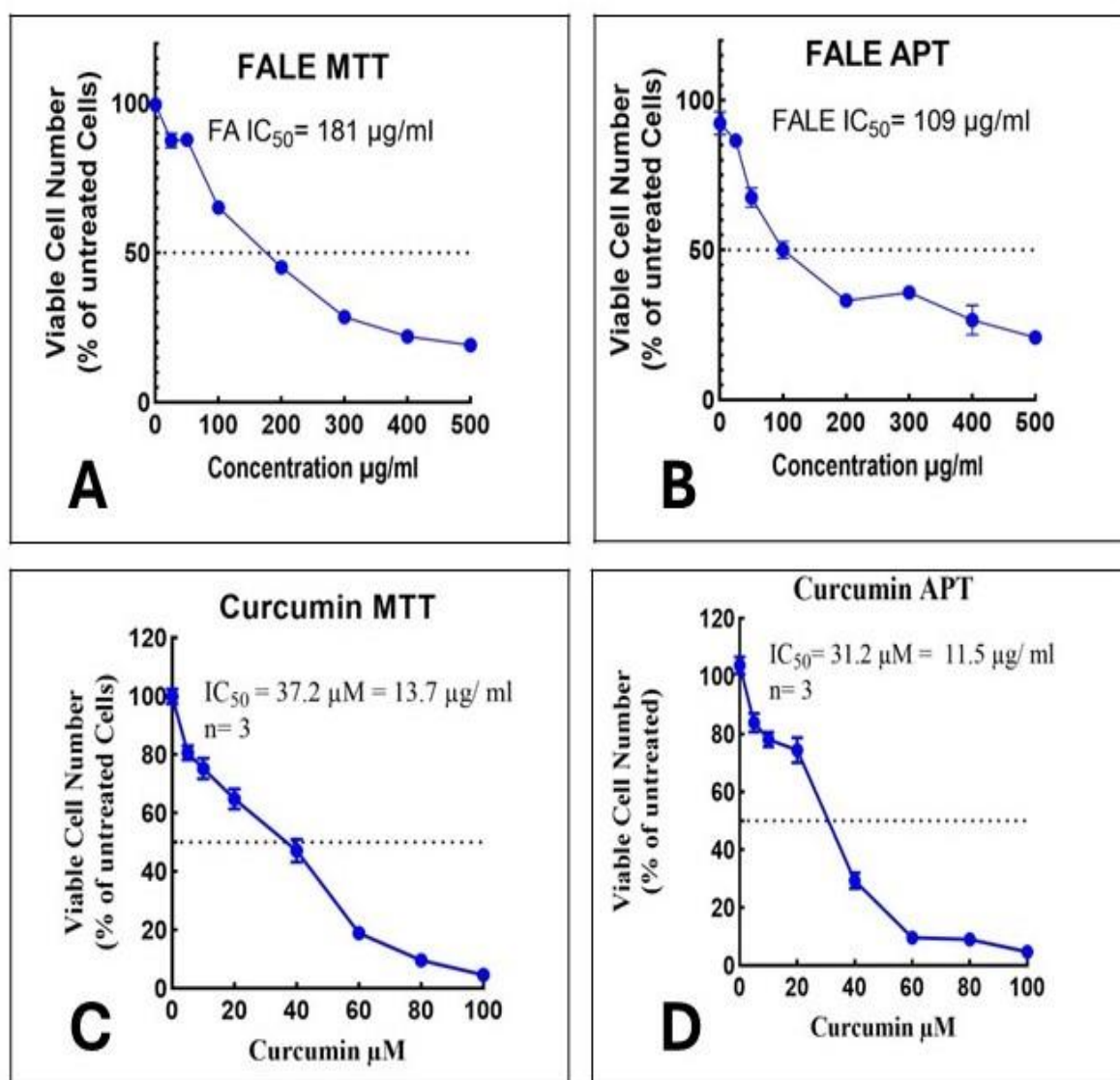


Figure 4: The percentage of MCF-7 cell death by MTT (3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium Bromide) assay and Acid phosphatase assay (APP) at different doses of FALE after 48h of incubation. **A.** MTT assay with FALE IC₅₀= 181 $\mu\text{g/ml}$ **B.** APT assay with FALE IC₅₀= 109 $\mu\text{g/ml}$ **C.** MTT assay with Curcumin IC₅₀ = 37.2 μM = 13.7 $\mu\text{g/ml}$ **D.** APT assay with Curcumin IC₅₀= 31.2 μM = 11.5 $\mu\text{g/ml}$. Curcumin is used as a standard drug. Each value represents a mean \pm SD (n = 3).

qPCR of targeted genes by FALE

Caspase-1, -3, -7, and -9 gene expression was evaluated in MCF-7 cells treated with FALE.

In cells treated with FALE, the $\Delta\Delta\text{Ct}$ mean of the Caspase-1 gene compared to GAPDH at 100 $\mu\text{g/ml}$ was 1.86 ± 0.95 , and at 300 $\mu\text{g/ml}$ was 2.59 ± 0.25 with no statistical difference between two doses and with the positive control Doxorubicin at 0.75 $\mu\text{g/ml}$. In contrast, the untreated MCF-7 cells exhibited a lower $\Delta\Delta\text{Ct}$ mean of the Caspase-1 gene, which was highly statistically significant. The $\Delta\Delta\text{Ct}$ mean of the Caspase-3 gene in cells treated with

FALE at 100 $\mu\text{g/ml}$ was 2.98 ± 0.98 , and at 300 $\mu\text{g/ml}$, it was 3.32 ± 0.57 with no statistical differences among both doses but revealed a significant difference with positive control (Figure 5A, Table 8). Untreated MCF-7 cells exhibited a much lower $\Delta\Delta\text{Ct}$ mean of the Caspase-3 gene, which was highly significant statistically (Figure 5B, Table 8). The Caspase-7 gene in cells treated with FALE at 100 $\mu\text{g/ml}$ was 2.32 ± 0.97 , and at 300 $\mu\text{g/ml}$ was 2.42 ± 0.25 and with of the Caspase-9 gene in cells treated at both doses were 2.63 ± 0.92 and 3.34 ± 0.44 respectively with statistical results similar to Caspase 3 (Figure 5C, 5D, Table 8).

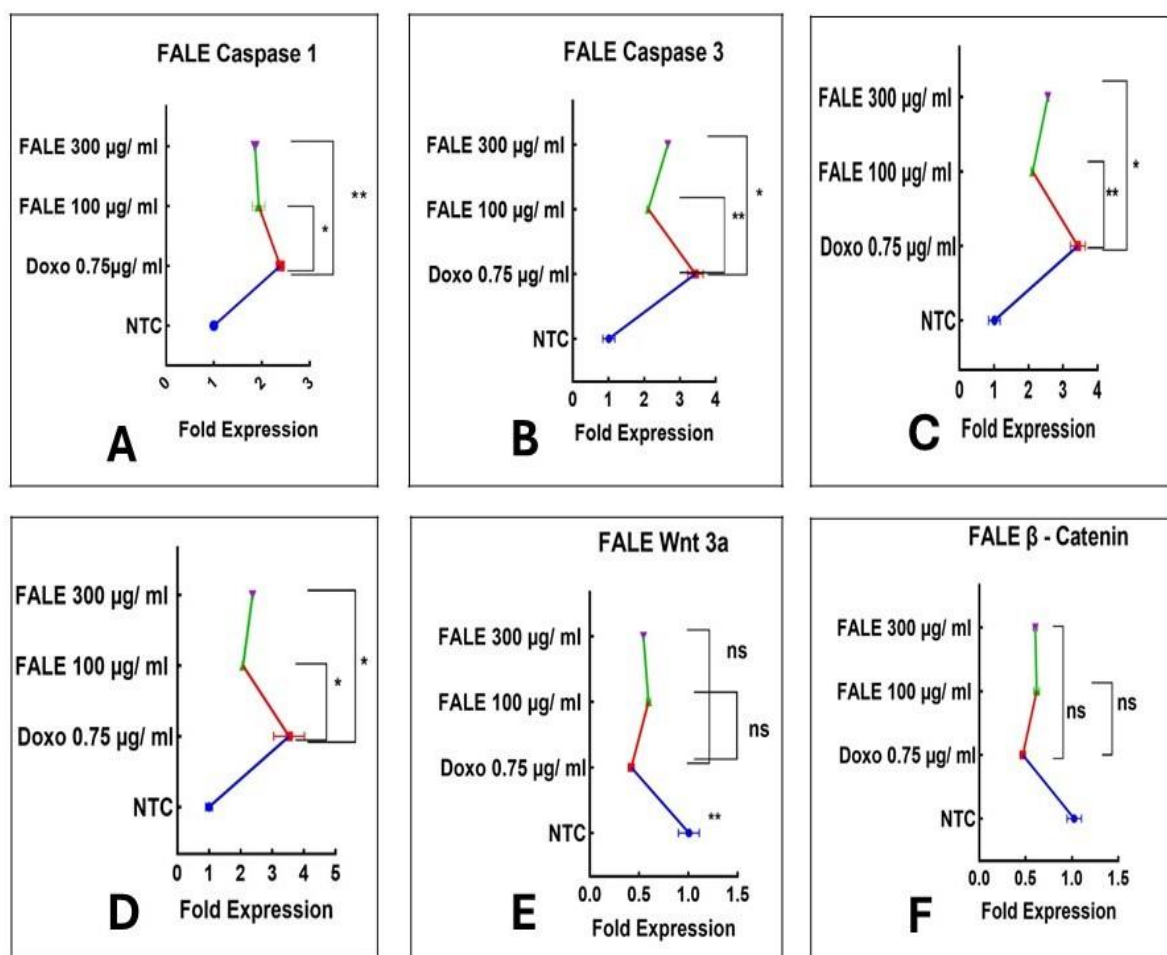


Figure 5 Comparative analysis in the expression of Caspases and Wnt genes compared to GAPDH in MCF-7 cells after 24 hr exposure with FALE. NTC: Non-treated cells, Doxo: Doxorubicin, a positive control, FALE: *Fagonia arabica* leaf extract. Comparing treated cells VS Non treated. All targeted genes showing Statistically significant difference of $p^{**} < 0.001$ and $p^{*} < 0.01$. A Casp- 1 gene B Casp- 3 gene C Casp- 7 gene D Casp- 9 gene E Wnt- 3a gene F β - catenin gene.

Table 8: Summary of fold expression of targeted genes by FALE (Real-time PCR) (n= 6).

Treatments	2 ^{^(- ΔΔ Cq)} Fold expression targeting Caspases and Wnt signaling genes					
	Casp 1	Casp 3	Casp 7	Casp 9	Wnt- 3a	β-Catenin
NTC	1.00 ± 0.04	1.01 ± 0.17	1.04 ± 0.41	1.00 ± 0.01	1.02 ± 0.28	1.00 ± 0.04
Doxorubicin 0.75 μg/ ml	2.38 ± 0.03	3.44 ± 0.22	3.26 ± 0.05	3.52 ± 0.48	0.14 ± 0.007	0.47 ± 0.02
FALE 100 μg/ ml	1.86 ± 0.95	2.98 ± 0.98	2.31 ± 0.97	2.64 ± 0.92	0.55 ± 0.93	0.41 ± 0.88
FALE 300 μg/ ml	2.59 ± 0.25	3.32 ± 0.57	2.43 ± 0.25	3.35 ± 0.44	0.35 ± 0.00	0.59 ± 0.03

The study observed a decrease in the expression of Wnt-3a and β-catenin genes following FALE compared to non-treated cells. The relative expression of the Wnt-3a gene at FALE 100 μg/ ml was 0.55 ± 0.94 , and at 300 μg/ ml was 0.34 ± 0.00 and of β-catenin gene at FALE 100 μg/ ml was 0.42 ± 0.88 , and at 300 μg/ ml was 0.58 ± 0.03 with no statistical difference. Similarly, comparison with Doxorubicin as the positive control revealed non-significant differences. In contrast, untreated MCF-7 cells exhibited a higher $\Delta\Delta C_t$ mean of the Wnt-3a gene and β-catenin gene at, which was statistically significant (Figure 5E, 5F Table 8). Overall, our results indicate a significant difference in the expression of all target genes in treated versus non-treated cells, suggesting upregulation of caspases and downregulation of Wnt-3a and β-catenin genes in MCF-7 cells upon treatment with FALE at two different doses. FALE shows potential as an effective anti-apoptotic and anticancer agent against MCF-7 cells through active gene modulation.

Discussion

Breast cancer incidence is on the rise globally due to factors such as urbanization, poor dietary habits, and the adoption of Western lifestyles, particularly in developing countries [26]. Despite advancements in chemotherapy, many cancer cells resist conventional treatments, prompting researchers to explore natural

alternatives with fewer side effects [27]. Polyphenols, abundant in plants, exhibit antioxidant properties that mitigate oxidative stress by scavenging free radicals and inhibiting lipid peroxidation [28]. Our investigation utilized crude plant extract, a common practice for assessing antioxidant activity and identifying bioactive components [29]. Crude extract antioxidant combinations can have complementary, antagonistic, or additive effects [29]. Methanolic extracts were chosen for research due to their higher yield. *Fagonia arabica*, a plant with traditional, ecological, therapeutic, and economic value, was evaluated for potential pharmaceutical use. The phytochemicals of *Fagonia arabica*, including primary metabolites that neutralize free radicals and secondary metabolites that prevent ROS formation, suggest its medicinal potential [30].

Plants contain various phytochemical families with therapeutic uses, including tannins, phenols, flavonoids, alkaloids, saponins, and other compounds. Tannins are polyphenolic substances with antiviral, antifungal, and antiparasitic properties, while flavonoids and phenols have antioxidant functions and are linked to cancer and inflammation. Alkaloids have anticancer action and are being studied for breast cancer treatment. The chemical makeup of medicinal herbs is influenced by species, growth conditions, age, and harvesting period [31]. Reactive oxygen species (ROS) are a therapeutic target in

tumor progression due to their fluctuating levels. Natural antioxidants like polyphenols, flavonoids, and carotenoids, found in medicinal plants, have been studied for suppressing ROS-producing enzymes, which are sometimes overexpressed in cancer. High intake of flavonoids has been linked to treating various diseases due to their antioxidant properties and electron transfer capacity [32]. Gallic acid and rutin were used as standards to quantify the total phenolic and flavonoid contents of FALE. According to our research, FALE has 50.02 ± 0.1 mg of rutin and 78.72 ± 0.5 mg of gallic acid/gram of extract, which closely agrees with data from earlier studies [33] signifies the antioxidant potential of FALE.

FTIR spectroscopy has become the method of choice in herbal medicine, often used more frequently than chromatographic techniques [34]. The capability of FTIR to concurrently separate several elements in comparable samples with a single experimental measurement is a powerful feature [35]. The identification of functional groups in bioactive compounds has been verified using FTIR research. It has been established that *Fagonia arabica* contains functional groups such as phenols, flavonoids, alkaloids, and terpenoids. These compounds may have pharmacological activity and potential functions in antioxidant and anticancer effects [21, 22, 36]. Numerous volatile and nonvolatile compounds can be found in plant extracts. In our investigation, 23 bioactive chemicals were identified using GC-MS analysis of FALE. Of these compounds, the two most abundant polyphenols in FALE were 2,3-Diphenyl-5-methoxybenzoic-1,4 (with a percentage abundance of 58.78%), reported for its anticancerous activity [37], and 3,7,11,15-Tetramethyl-2-hexadec (with a percentage abundance of 11.81%), which has been linked to anticancer and antibacterial activities [38]. To provide similar results,

our study was mainly created to evaluate the cytotoxic potential of plant extract with that of conventional medications. Several preliminary studies were performed to confirm the antioxidant activity of *Fagonia arabica* leaf extract (FALE) and assess its potential for cytotoxicity. A practical method for assessing test sample antioxidant activity *in vitro* is cytotoxicity analysis. Using Curcumin as a conventional medication, we got equivalent outcomes in our trial. A study [39] determined that 100 $\mu\text{g}/\text{ml}$ was the highest effective dose, which was inconsistent with the IC_{50} value of 181 $\mu\text{g}/\text{ml}$ obtained from the MTT experiment for FALE. On the other hand, the Curcumin MTT assay produced an IC_{50} value of 13.7 $\mu\text{g}/\text{ml}$, which aligns the reported findings [40]. Furthermore, the FALE Acid Phosphatase Assay gave an IC_{50} value of 109 $\mu\text{g}/\text{ml}$, which has not been published because no assay of this kind has been carried out with *Fagonia arabica*. The findings [41] associated with the Acid Phosphatase Assay for Curcumin, which showed an IC_{50} value of 11.5 $\mu\text{g}/\text{ml}$.

qPCR was utilized to examine the fold expression of caspase-1, -3, -7, and -9, as well as the Wnt-3a and β -catenin genes by using FALE at 100 $\mu\text{g}/\text{ml}$ and 300 $\mu\text{g}/\text{ml}$. Our findings showed that caspases were upregulated while the Wnt-3a and β -catenin genes were downregulated in a statistically significant way ($P < 0.001$). These results line up with comparable published research. Researcher showed that Casp-1 performs a caspase-1-dependent process of cell death triggered by microbial infections known as pyroapoptosis. Xanthoangelol (XAG), isolated from *Angelica keiskei*, has shown a drug-dependent increase in Casp-1 activity in Hep-G 2 cell lines [42]. The effect of XAG on Casp-1-dependent pyroptosis, indicating the therapeutic potential of natural compounds [43]. Furthermore, the real-time PCR study of *Zea mays* cotton silk extract, discovered a substantial elevation in the fold expression

of Casp-3 and -9, showing enhanced apoptosis ($P < 0.05$) [44]. Emodin, an anthraquinone derivative separated from *Rheum palmatum L.*, a Chinese herb, caused a significant ($P < 0.05$) increase in Caspase-9 and -3 expression in a dose-dependent manner, causing apoptosis through death receptor and mitochondrial pathways [45]. Comparing to control samples, Emodin treatment significantly increased Caspase-3 and -9-fold expression ($P < 0.001$), with decreased expression of Wnt-3a and β -catenin cancer cell lines. Another investigation for the function of miR-195 in colon cancer revealed significant results for both cell viability and β -catenin levels [46]. Doxorubicin treatment proved cytotoxic to MCF-7 cells, to reduce the dosage and its adverse effects, resveratrol was investigated on enhancing the sensitivity of MCF-7 breast cancer cells to doxorubicin [47]. These compounds are receiving more attention in the drug development process due to their anticancer qualities. Here, we found that *Fagonia arabica* significantly downregulated β -catenin and Wnt-3a in MCF-7 cells.

Conclusion

The leaves of *Fagonia arabica* are abundant in polyphenols according to phytochemical studies. Strong antioxidant activity was shown by the greatest amounts of gallic acid and rutin in the methanolic fraction. FTIR and GC-MS spectra provided objective evidence of the diverse chemical constituents present in this complex system, validating its traditional and herbal medicinal uses. Further research may uncover new bioactive compounds in *Fagonia arabica*. *Fagonia arabica* could be a promising natural source of antioxidants for breast cancer treatment. Our study demonstrated the upregulation of caspase-1, -3, -7, and -9, the proapoptotic marker genes, and the downregulation of Wnt-3a and β -catenin, indicating apoptotic cell death in MCF-7 cells. Therefore,

Fagonia arabica holds potential as a foundation for developing novel lead structures in cancer drug strategies. However, further assessment of its bioactive compounds using techniques such as HPLC and NMR and antioxidant activities in animal models is warranted. Rigorous, well-designed clinical trials are essential to validate the efficacy and safety of natural products in treating breast cancer. For more conclusive proof, future studies should concentrate on large-scale experiments. Our findings may help treat breast cancer with additional validation.

Acknowledgment: The authors would also like to thank the Higher Education Commission (HEC) for funding of the study through the IRSIP program.

Author contributions: All authors contributed to the study's conception and design. The first draft of the manuscript was written by Ambreen Sher and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

Conflict of interest: The authors declare no conflict of interest.

Data availability: All data is presented in the manuscript.

Ethics approval: The study was approved by the Institutional Review Board (or Ethics Committee) of the Department of Biological Sciences, International Islamic University, Islamabad 44000, Pakistan, and was given an ethical approval number: No. IIUI/ORIC/Bioethics/110-81. The MCF-7 human breast cancer cell lines used in this study were obtained from the European Collection of Authenticated Cell Cultures (ECACC).

References

1. Koni M, Pinnarò V & Brizzi M F. The

- Wnt* signalling pathway: a tailored target in cancer. *International Journal of Molecular Sciences*. 2020; 21(20):7697.
2. Mukherjee A, Stathos M E, Varner C, Arsiwala A, Frey S, Hu Y, & Kane R S. One-pot synthesis of heterodimeric agonists that activate the canonical *Wnt* signaling pathway. *Chemical Communications*. 2020; 56(25):3685-3688.
 3. Jackstadt R, Hodder MC, & Sansom OJ. *WNT* and β -Catenin in Cancer: Genes and Therapy. *Annual Review of Cancer Biology*. 2020; 4:177-196.
 4. Merikhian P, Eisavand MR & Farahmand L. Triple-negative breast cancer: Understanding *Wnt* signaling in drug resistance. *Cancer Cell International*. 2021; 21(1):1-8.
 5. Casella D, Palumbo P, Sandroni S, Caponi C, Littori F, Capuano F, ... & Cuomo R. Positive ROS (Reactive Oxygen Species) Modulator Engineered Device Support Skin Treatment in Locally Advanced Breast Cancer (LABC) Enhancing Patient Quality of Life. *Journal of Clinical Medicine*. 2022; 11(1):126.
 6. Ahmed HM, Nabavi S, & Behzad S. Herbal drugs and natural products in the light of nanotechnology and nanomedicine for developing drug formulations. *Mini Reviews in Medicinal Chemistry*. 2021;21(3): 302-313.
 7. Lepourcelet M, Chen YNP, France DS, Wang H, Crews P, Petersen F, ... & Shivdasani RA. Small-molecule antagonists of the oncogenic Tcf/ β -catenin protein complex. *Cancer cell*. 2004; 5(1):91-102.
 8. Gajos-Michniewicz A, Czyz M. Therapeutic Potential of Natural Compounds to Modulate WNT/ β -Catenin Signaling in Cancer: Current State of Art and Challenges. *International Journal of Molecular Sciences*. 2024; 25(23):12804.
 9. Soni V, Nagar A, Bardiya R, Mara J, Von Suskil L, Rose S & Sonawane C. A Concise Review of Prodigious Salinomycin and Its Derivatives Effective in Treatment of Breast Cancer. *International Journal of Translational Medicine*. 2023; 3(2):217-245.
 10. Prasad CP, Rath G, Mathur S, Bhatnagar D, & Ralhan R. Potent growth suppressive activity of curcumin in human breast cancer cells: Modulation of *Wnt*/ β -catenin signaling. *Chemico-biological interactions*. 2009; 181(2):263-271.
 11. Ali MZ, Mehmood MH, Saleem M, Akash MSH & Malik A. (2021). Pharmacological evaluation of *Euphorbia hirta*, *Fagonia indica* and *Capparis decidua* in hypertension through in-vivo and in vitro-assays. *Heliyon*. 2021; 7(10).
 12. Mansoor S, Shahid S, Javed M, Saad M, Iqbal S, Alsaab HO, ... & Fatima A. Green synthesis of a MnO-GO-Ag nanocomposite using leaf extract of *Fagonia arabica* and its antioxidant and anti-inflammatory performance. *Nano-Structures & Nano-Objects*. 2022; 29, 100835.
 13. Malavika PS, Singh V, & Kumar Y. The chemistry and pharmacology of *Fagonia* genus: a review. *IJSDR*. 2021; 6(7):332-343.
 14. Kashif M, Abbas A, Nasir A, Rehman AU, Rehman E, Jamal MA, Kazi M. Exploring the Antiproliferative and Antioxidant Effects of *Fagonia cretica* Thereof: An In Vitro and In Vivo Evaluation. *Microscopy Research and Technique*. 2025 Feb 24.
 15. Thanh DT, Tan MT, Thu NTM, Trinh PNP, Thuong PTH, Tuyet PTG & Hieu TT. Phytochemical Composition, Antioxidant, Anti-*Helicobacter pylori*, and Enzyme Inhibitory Evaluations of *Cleistocalyx operculatus* Flower Bud and Leaf Fractions. *BioTech*. 2024; 13(4):42.

16. Kanwal I, Fatima N, Wazir A, Khan M, Zaheer M, Masroor D, & Syed S. *Fagonia Arabica* Linn, a miraculous medicinal plant with diminutive scientific data but hefty potential. *RADS Journal of Pharmacy and Pharmaceutical Sciences*. 2021; 9(3):185-189.
17. Badawy SA, Hassan AR, Abu Bakr MS, Mohammed AE. UPLC-qTOF-MS/MS profiling of phenolic compounds in *Fagonia arabica* L. and evaluation of their cholinesterase inhibition potential through in-vitro and in-silico approaches. *Scientific Reports*. 2025;15(1):5244.
18. Javed H, Tabassum S, Erum S, Murtaza I, Muhammad A, Amin F, & Nisar MF. Screening and characterization of selected drugs having antibacterial potential. *Pakistan journal of pharmaceutical sciences*. 2018; 31(3).
19. Batool R, Khan MR, Sajid M, Ali S, & Zahra Z. Estimation of phytochemical constituents and in vitro antioxidant potencies of *Brachychiton populneus* (Schott & Endl.) R. Br. *BMC chemistry*. 2019; 13:1-15.
20. Sher A, Tabassum S, Wallace HM, Khan A, Karim AM, Gul S, Kang SC. In vitro analysis of cytotoxic activities of *Monotheca buxifolia* targeting WNT/ β -catenin genes in breast cancer cells. *Plants*. 2023;12(5):1147.
21. Ayaz M, Junaid M, Ullah F, Sadiq A, Shahid M, Ahmad W, ... & Syed NIH. GC-MS analysis and gastroprotective evaluations of crude extracts, isolated saponins, and essential oil from *Polygonum hydropiper* L. *Frontiers in Chemistry*. 2017; 5:58.
22. Nazir M, Khan SA, & Bhatti MK. The constituents of unsaponifiable from *Monotheca buxifolia* seed oil. *Fette, Seifen, Anstrichmittel*. 1986; 88(7):266-268.
23. Akinyele O, & Wallace HM. Characterising the response of human breast cancer cells to polyamine modulation. *Biomolecules*. 2021; 11(5):743.
24. Rio DC, Ares M, Hannon GJ, & Nilsen TW. Purification of RNA using TRIzol (TRI reagent). *Cold Spring Harbor Protocols*, 2010;(6): pdb-prot5439.
25. Livak KJ, & Schmittgen, TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2- $\Delta\Delta$ CT method. *Methods*. 2002; 25(4):402-408.
26. Kashyap D, Pal D, Sharma R, Garg VK, Goel N, Koundal D, Zaguia A, Koundal S, Belay A. [Retracted] Global increase in breast cancer incidence: risk factors and preventive measures. *BioMed research international*. 2022;(1):9605439.
27. Huang M, Lu JJ & Ding J. Natural products in cancer therapy: Past, present and future. *Natural products and bioprospecting*. 2021; 11:5-13.
28. Singh GK. Medicinal Properties and Therapeutic Potential of *Fagonia arabica* Linn: A Comprehensive Review. *Asian Journal of Pharmaceutics (AJP)*. 2024;18(3).
29. Arif Y, Bajguz A, & Hayat S. *Moringa oleifera* extract as a natural plant biostimulant. *Journal of Plant Growth Regulation*. 2022; 1-16.
30. Marwat SK, Rehman, FU, Usman K, Khakwani AZ, Ghulam S, Anwar N & Sadiq M. Medico-ethnobotanical studies of edible wild fruit plants species from the flora of north western Pakistan (DI Khan district). *J Med Plants Res*. 2011; 5(16):3679-3686.
31. Chen D, Ma Y, Guo Z, Liu L, Yang Y, Wang Y, ... & Yang W. Two natural alkaloids synergistically induce apoptosis in breast cancer cells by inhibiting STAT3 activation. *Molecules*. 2020; 25(1):216.
32. Saikolappan S, Kumar B, Shishodia G, Koul S, & Koul HK. Reactive oxygen species and cancer: A complex interaction. *Cancer letters*. 2019; 452:132-143.

33. Khan I, Ali JS, Ul-Haq I, & Zia M. Biological and phytochemicals properties of *Monotheca buxifolia*: an unexplored medicinal plant. *Pharmaceutical Chemistry Journal*. 2020; 54:293-301.
34. Zou Y, Li Y, Hao W, Hu X, & Ma G. Parallel β -sheet fibril and antiparallel β -sheet oligomer: New insights into amyloid formation of hen egg white lysozyme under heat and acidic condition from FTIR spectroscopy. *The Journal of Physical Chemistry B*, 2013;117(15):4003-4013.
35. Moro MK, dos Santos FD, Folli GS, Romão W, Filgueiras PR. A review of chemometrics models to predict crude oil properties from nuclear magnetic resonance and infrared spectroscopy. *Fuel*. 2021; 303:121283.
36. Nawaz F, Ahmad S, Naheed S, Ahmad Z, & Mehmmod T. Phytochemical screening by FTIR spectroscopic analysis of leaf extracts of *Monotheca Buxifolia*. *University of Wah Journal of Science and Technology (UWJST)*. 2019; 3:15-22.
37. Dilshad R, Khan KUR, Saeed L, Sherif A E, Ahmad S, Ovatlarnporn C, ... & Mukhtar I. Chemical composition and biological evaluation of *Typha domingensis* pers. to ameliorate health pathologies: In vitro and in silico approaches. *BioMed Research International*, 2022.
38. Ismail GA, Gheda SF, Abo-shady AM, & Abdel-karim OH. In vitro potential activity of some seaweeds as antioxidants and inhibitors of diabetic enzymes. *Food Science and Technology*. 2019; 40:681-691.
39. Satpute R, Bhattacharya R, Kashyap RS, Purohit HJ, Deopujari, JY, Taori, GM, & Dagainawala HF (2012). Antioxidant potential of *Fagonia arabica* against the chemical ischemia-induced in PC12 cells. *Iranian journal of pharmaceutical research: IJPR*, 11(1):303.
40. Zhou QM, Wang XF, Liu XJ, Zhang H, Lu YY & Su SB. Curcumin enhanced antiproliferative effect of mitomycin C in human breast cancer MCF-7 cells in vitro and in vivo. *Acta Pharmacologica Sinica*. 2011;32(11):1402-1410.
41. Ravish I, & Raghav N. Curcumin as inhibitor of mammalian cathepsin B, cathepsin H, acid phosphatase and alkaline phosphatase: a correlation with pharmacological activities. *Medicinal Chemistry Research*. 2014; 23:2847-2855.
42. Wang C, Wang L, Huang C, Liu Y, Liu J, Kuang H, ... & Fan R. Involvement of NLRP3/Caspase-1/GSDMD-dependent pyroptosis in BPA-Induced apoptosis of human neuroblastoma cells. *Biochemical Pharmacology*. 2022; 200:115042.
43. Pang X, Gao X, Liu F, Jiang Y, Wang M, Li Q, & Li Z. Xanthoangelol modulates Caspase-1-dependent pyroptotic death among hepatocellular carcinoma cells with high expression of GSDMD. *Journal of Functional Foods*. 2021; 84:104577.
44. Al-Oqail MM, Al-Sheddi ES, Farshori NN, Al-Massarani SM, Al-Turki EA, Ahmad J, ... & Siddiqui M. Corn silk (*Zea mays* L.) induced apoptosis in human breast cancer (MCF-7) cells via the ROS-mediated mitochondrial pathway. *Oxidative Medicine and Cellular Longevity*, 2019.
45. Yaoxian W, Hui Y, Yunyan Z, Yanqin L, Xin G & Xiaoke W. Emodin induces apoptosis of human cervical cancer hela cells via intrinsic mitochondrial and extrinsic death receptor pathway. *Cancer cell international*. 2013;13(1):1-8.
46. Li B, Wang S & Wang S. MiR-195 suppresses colon cancer proliferation and metastasis by targeting *WNT3A*. *Molecular Genetics and Genomics*. 2018; 293(5):1245-1253.
47. Osman AMM, Bayoumi HM, Al-Harathi SE, Damanhoury ZA, & ElShal MF.

- Modulation of doxorubicin cytotoxicity by resveratrol in a human breast cancer cell line. *Cancer cell international*. 2012; 12(1):1-8.
48. Shakeel A, Sajid L, Ahmad MA, Zafar S. Investigating Accumulative Effect of Fluoxetine and Fagonia indica as an Anti-Stress Therapy (Doctoral dissertation, Atta Ur Rahman School of Applied Biosciences (ASAB), NUST).
 49. Saharan V, Mehrotra A, Khatik R, Rawal P, Sharma SS, Pal A. Synthesis of chitosan-based nanoparticles and their in vitro evaluation against phytopathogenic fungi. *International journal of biological macromolecules*. 2013; 62: 677-83.
 50. Nivedita S, Joseph S. Optimization of process parameters using response surface methodology for PCL based biodegradable composite membrane for water purification. *Arabian Journal for Science and Engineering*. 2020; 45: 7347-60.
 51. Pasela BR, Gammag RP, Castillo AP, Tumacder DV, Taaca KL. Straightforward Synthesis of Cost-Effective, Flexible, and Hydrophobic Polyaniline-Chitosan (PAni-Cs) Films Produced with Lactic Acid as Solvent. *In Materials Science Forum*. 2021; 1041: 29-35.
 52. Jamaluddin F, Mohameda S, Lajis MN. Hypoglycaemic effect of stigmast-4-en-3-one, from Parkia speciosa empty pods. *Food Chemistry*. 199; 54(1):9-13.
 53. Ismail GA, Gheda SF, Abo-Shady AM, Abdel-Karim OH. In vitro potential activity of some seaweeds as antioxidants and inhibitors of diabetic enzymes. *Food Science and Technology*. 2019; 40: 681-91.
 54. Khan MS, Uzair M, Hanif M, Jabeen M, Chaudhary BA. Anti-inflammatory potential of spectroscopically analyzed trans-13-octadecenoic acid of Yucca elephantipes Regel roots: in-vitro and in-vivo analysis. *Pakistan Journal of Pharmaceutical Sciences*. 2022; 35(6).
 55. Peng WX, Xue Q, Wu FJ, Zhang X, Zhang ZF. Analysis on health risk of volatile organic compounds of smoked bamboo biomaterial by TD-GC-MS. *Key Engineering Materials*. 2011; 467:1697-701.
 56. Dilshad R, Khan KU, Saeed L, Sherif AE, Ahmad S, Ovatlarnporn C, Nasim J, Hussain M, Ghalloo BA, Basit A, Mukhtar I. [Retracted] Chemical Composition and Biological Evaluation of Typha domingensis Pers. to Ameliorate Health Pathologies: In Vitro and In Silico Approaches. *BioMed Research International*. 2022; 2022(1):8010395.
 57. Uthpala TG, Munasinghe HH, Peiris LD, Navaratne SB. Evaluation of antimicrobial potential and phytochemicals in Acemella (A. oleracea) flower pod extracts subjected to different drying techniques. *Journal of Food Processing and Preservation*. 2021; 45(6):15570.
 58. Maher T, Kabbashi NA, Mirghani ME, Alam MZ, Daddiouaissa D, Abdulhafiz F, Reduan MF, Omran JI, Abdul Razab MK, Mohammed A. Optimization of ultrasound-assisted extraction of bioactive compounds from Acacia seyal gum using response surface methodology and their chemical content identification by Raman, FTIR, and GC-TOFMS. *Antioxidants*. 2021;10(10):1612.
 59. Luhata LP, Yoshida Y, Usuki T. Natural products from Odontonema strictum promote neurite outgrowth in neuronal PC12 cells. *Bioorganic Chemistry*. 2024; 147:107389.
 60. Ren F, Zhang Y, Qin Y, Shang J, Wang Y, Wei P, Guo J, Jia H, Zhao T. Taraxasterol prompted the anti-tumor effect in mice burden hepatocellular carcinoma by regulating T lymphocytes. *Cell Death Discovery*. 2022 ;8(1):264.